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Systematic Review for Phytotherapy in Streptococcus Mutans

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Abstract

Streptococcus mutans is one of the bacteria that are a major cause of tooth decay. Streptococcus mutans plays a special role in plaque formation and tooth decay. *S. mutans*, which is a gram-positive cocci and encapsulated bacterium, produces acid and is considered as an extracellular glucan polymer. Medicinal plants are of special value and importance in securing health status at the society level in terms of both treatment and prevention of human diseases and are as old as human history. The present review study reports on medicinal plants that are used as antibacterials against *S. mutans*. In this study, the search process was done using keywords, such as bacteria, Streptococcus mutans, and medicinal plants. Databases such as ISI Web of Science, PubMed, Scopus, Google Scholar and Science Direct were searched and the relevant articles were used for reviewing purposes. Based on the results, there are 48 medicinal plants with anti-*S. mutans* effects.

Keywords: Anti-Bacterial, Streptococcus mutans, Medicinal plants

INTRODUCTION

Streptococcus mutans and *Streptococcus sangui* are two of the bacteria considered as major cause of tooth decay [1-5]. Some species such as Streptococcus mutans streptococci play special role in plaque formation and tooth decay [6]. *Streptococcus mutans*, which is a gram-positive cocci and encapsulated bacterium, produces acid and is considered as an extracellular glucan polymer [7]. When mutans population reaches 50% of the total population of bacteria, it then can cause tooth decay [7]. Streptococcus mutans binds to the teeth by creating homo-polymers glucan from sucrose via glucosyltransferase [8].

Medicinal plants are of special value and importance in securing health status at the society level in terms of both treatment and prevention [9-17] of human diseases. These are parts of natural resources (18-22) and are as old as human history [23-25] and have been one of the most important sources of human food and medicine over generations [26-35]. From a historical viewpoint, plants have an important role in community development and extensive researches have been conducted to find herbal

medicinal products and natural materials throughout history [23, 36-48]. Due to safety of use in comparison to conventional drugs, and antibiotic resistance, people prefer to use them and researchers conduct a lot of researches on them [49-58]. The present review study reports the medicinal plants that are used as antibacterial against *Streptococcus mutans*.

METHODS

In this study, the search process was done using keywords, such as bacteria, Streptococcus mutans, and medicinal plants. Databases such as ISI Web of Science, PubMed, Scopus, Google Scholar and Science Direct were searched and the relevant articles were used for reviewing purposes.

RESULTS

Based on the results, 48 medicinal plants are used as antimicrobials against *Streptococcus mutans*. Medicinal herbs with anti-*streptococcus mutans* effects along with their additional information are shown in Table 1.

Table1. Antimicrobial medicinal herbs against Streptococcus mutans along with their Farsi name, scientific name, family name

family name								
Scientific Name	Part of plant	Family Name	Origin of plant	Country of study	Bacteria	Result	Ref.	
Psidium guajava	leaf	Myrtaceae	India	India	human dental Streptooccus mutans	Diameter of inhibition zone induced by ethyl acetate extract (5 and 2.5 mg/discs) of the plant was 20 mm and 18 mm, respectively. Minimum inhibitory concentration of less than 0.076 (MIC <0.076 mg / mL) was reported.	[59]	
Terminalia chebula	fruit	Combretaceae	India	India	human dentalStreptococcus mutans	Diameter of inhibition zone induced by ethyl acetate extract (5 and 2.5 mg/discs) of the plant was 16 mm and 14 mm, respectively. Minimum inhibitory concentration of less than 0.076 (MIC <0.076 mg / mL) was reported.	[59]	
Mimus opselengi	stick	Sapotaceae	India	India	human dentalStreptococcus mutans	Diameter of inhibition zone induced by ethyl acetate extract (5 and 2.5 mg/discs) of the plant was 16 mm and 14 mm, respectively.	[59]	
Achyranthes aspera	Root	amaranthaceae	India	India	human dentalStreptococcus mutans	Diameter of inhibition zone induced by ethyl acetate extract (5 and 2.5 mg/discs) of the plant was 18 mm and 12 mm, respectively.	[59]	
Polygonumcuspidatum	root	Polygonaceae	Korea	Republic of Korea	S. mutans KCTC 3298	The antibacterial effect of methanol extract of the plant was proved at MIC = $0.5-4$ mg / ml and MBC 2 to 4 times higher than the MIC.	[60]	
CINNAMOMUM CASSIA	bark	Lauraceae	Pakista n	Pakistan	Streptococcus mutans	The inhibition zone diameter of the plant was respectively 29, 27.15, 18, 19, 32, 19, 27 and 15 for the corresponding bacteria.	[61]	
Mikanialaevigata	aerial parts	Asteraceae	Brazil	Brazil	culture collection and clinical isolates of Streptococcus mutans	Hexane extract of the plant with MIC between 12.5 and 400µg / ml and MBC between 25 and 400µg / ml proved to inhibit the bacterial growth. The bacterium ability to bind to the glass plate was also inhibited.	[61]	
Mikania glomerata	aerial parts	Asteraceae	Brazil	Brazil	culture collection and clinical isolates of Streptococcus mutans	Hexane extract of the plant with MIC between 12.5 and 400µg / ml and MBC between 25 and 400µg / ml proved to inhibit the bacterial growth. The bacterium ability to bind to the glass plate was also inhibited.	[62]	
Glycyrrhiza glabra	rhizom es	Fabaceae	Thailan d	Thailand	Streptococcus mutans (DMST18777)	It is proved that the ethanol extract inhibits the bacterial growth at a concentration of $20\mu g$ / ml with inhibition zone diameter of 15 mm and MIC<12.5 μg / ml and MBC = $25\mu g$ / ml. However, the inhibition zone diameter of 15 μg / ml erythromycin was 40 mm.	[63]	
Terminalia bellirica	fruit	Combretaceae	Thailan d	Thailand	Streptococcus mutans (DMST18777)	It is proved that the ethanol extract inhibits the bacterial growth at a concentration of $20\mu g$ / ml with inhibition zone diameter of 10 mm and MIC<12.5 μg / ml and MBC = $25\mu g$ / ml. However, the inhibition zone diameter of 15 μg / ml erythromycin was 40 mm.	[63]	
Syzygium aromaticum	Flower bud	Myrta ceae	Thailan d	Thailand	Streptococcus mutans (DMST18777)	It is proved that the ethanol extract inhibits the bacterial growth at a concentration of $20\mu g$ / ml with inhibition zone diameter of 14 mm and MIC<12.5 μg / ml and MBC = $50\mu g$ / ml proved. However, the inhibition zone diameter of 15 μg / ml erythromycin was 40 mm.	[63]	

Scientific Name	Part of plant	Family Name	Origin of plant	Country of study	Bacteria	Result	Ref.
Rosmarinus officinalis	Leaves	Lamiaceae	Taiwan	Taiwan	Streptococcus mutans (BCRC10793)	The methanolic plant extract inhibits the growth of bacterium at MIC = 4 mg / ml.	[64]
Haematoxyl onbrasiletto	Stem bark	Leguminosae	Mexico	Mexico	Streptococcus mutans (ATCC 10449)	Antibacterial property of aqueous extract of this plant was proved at MIC = $10.5\mu g / ml$ and MBC = $125\mu g / ml$. This is despite the fact that chlorhexidine, as control, has MIC = $1.2\mu g / ml$ and MBC = $2.7\mu g / ml$.	[65]
Punica granatum	Pericar p	Punicaceae	Mexico	Mexico	Streptococcus mutans (ATCC 10449)	Antibacterial property of aqueous extract of this plant was proved at MIC = $12.5\mu g / ml$ and MBC = $125\mu g / ml$. This is despite the fact that chlorhexidine, as control, has MIC = $1.2\mu g / ml$ and MBC = $2.7\mu g / ml$.	[65]
Iostephaneheterophylla	Roots	Compositae	Mexico	Mexico	Streptococcus mutans (ATCC 10449)	Antibacterial property of aqueous extract of this plant was proved at MIC = $67.5\mu g / ml$ and MBC = $125\mu g / ml$. This is despite the fact that chlorhexidine, as control, has MIC = $1.2\mu g / ml$ and MBC = $2.7\mu g / ml$.	[65]
Bursera simaruba	Stem bark	Burseraceae	Mexico	Mexico	Streptococcus mutans (ATCC 10449)	Antibacterial property of aqueous extract of this plant was proved at MIC = $100\mu g / ml$ and MBC = $500\mu g / ml$. This is despite the fact that chlorhexidine, as control, has MIC = $1.2\mu g / ml$ and MBC = $2.7\mu g / ml$.	[65]
Cedrela odorata	Seeds	Meliaceae	Mexico	Mexico	Streptococcus mutans (ATCC 10449)	Antibacterial property of aqueous extract of this plant was proved at MIC = $32.5\mu g / ml$ and MBC = $250\mu g / ml$. This is despite the fact that chlorhexidine, as control, has MIC = $1.2\mu g / ml$ and MBC = $2.7\mu g / ml$.	[65]
Rhus standleyi	Aerial parts	Anacardiaceae	Mexico	Mexico	Streptococcus mutans (ATCC 10449)	Antibacterial property of aqueous extract of this plant was proved at MIC = $32.5\mu g$ / ml and MBC = $125\mu g$ / ml. This is despite the fact that chlorhexidine, as control, has MIC = $1.2\mu g$ / ml and MBC = $2.7\mu g$ / ml.	[65]
Syzygium aromaticum	Flower bud	Myrtaceae	Mexico	Mexico	Streptococcus mutans (ATCC 10449)	Antibacterial property of aqueous extract of this plant was proved at MIC = $25\mu g / ml$ and MBC = $250\mu g / ml$. This is despite the fact that chlorhexidine, as control, has MIC = $1.2\mu g / ml$ and MBC = $2.7\mu g / ml$.	[65]
Helichrysum odoratissimum	Aerial part	ASTERACEAE	Uganda	Uganda	Streptococcusmutans (CCUG6519 T)	Anti-bacterial property of the hexane extract of this plant was proved at a minimum inhibitory concentration of 0.25 mg / ml. However, the minimum inhibitory concentration of doxycycline, as control, was reported 1.56 × 10-4 mg / ml.	[66]
Lantana trifolia	Leaves	VERBENACEA E	Uganda	Uganda	Streptococcusmutans (CCUG6519 T)	Anti-bacterial property of hexane and methanol extracts of this plant was proved at minimum inhibitory concentration of I mg / ml. However, the minimum inhibitory concentration of doxycycline, as control, was reported to be $1.56 \times$ 10-4 mg / ml were.	[66]

Scientific Name	Part of plant	Family Name	Origin of plant	Country of study	Bacteria	Result	Ref.
Kaempferiapandurata	rhizom e	Zingiberaceae	Indone	South Korea	Streptococcus mutans ATCC 25175	Anti-bacterial property of isopanduratin A compound in this plant was proved at MIC = 4 mg / ml and MBC = 8 mg / ml. This is despite the fact that chlorhexidine and vancomycin, as antibiotics, have MIC = 1 mg / ml and MBC = 2 mg / ml. On the other hand, 20 mg /ml isopanduratin A deactivates the bacteria within 1 minute.	[67]
Rheedia brasiliensis	fruits		Brazil	Brazil	Streptococcus mutans UA159	The antibacterial property of hexane extract of the fruit peels was proved at MIC = $12.5-25\mu g$ / ml or MBC = $25-50 \mu g$ / ml. Hexane extract of the fruit seeds of this plant also has MIC = $12.5-25\mu g$ / ml and MBC = $50-100\mu g$ / ml. Anti-bacterial property of the active ingredient of the plant (7- epiclusianone) was proved at MIC = $1.25-2.5\mu g$ / ml and MBC = $10-20\mu g$ / ml.	[68]
Stevia rebaudiana	leaves	Asteraceae	India	India	S. mutans (MTCC 497)	Antibacterial property of methanol extract of this plant was proved at a zone diameter of 11 mm. However, inhibition zone diameter of the antibiotic control was 10 mm.	[69]
Cinnamomumspp.		Lauraceae	India	India	Streptococcus mutans (MTCC 497)	Anti-bacterial property of the active ingredient of Cinnamaldehyde was proved as discs with concentrations of 20mM (diameter of inhibition zone of 30 mm), 80mM (diameter of inhibition zone of32mm) and 120mM (diameter of inhibition zone of 30mm). Inhibition zone diameter of was 16 mm was obtained for the active ingredient eugenol disc with a concentration of 120 mM. However, inhibition zone diameter of 18mm was obtaned for vancomycin (antibiotic 30 mcg disc) ,as a positive control.	[70]
Salvadorapersica	stems	Salvadoraceae	IRAQ	IRAQ	Clinical isolates of Streptococcus mutans	Antibacterial property of the aqueous extract of this plant was proved at a concentration of 200 mg / ml, with a inhibition zone diameter of 19.3 mm and MIC = 1.56 mg / ml. This is despite the fact that streptomycin as control has a inhibition zone diameter of 19.5 mm and MIC = 0.097 mg/ml.	[71]
Spilanthescalva	Floral parts	Asteraceae	India	India	Clinical isolates of Streptococcus mutans	Antibacterial property of aqueous extract of this plant was proved with the inhibition zone diameter of 11.6 mm. However, the inhibition zone diameter of following antibiotic controls was as follows: ampicillin (10 mm), nalidixic acid (11 mm), nitrofurantoin (10.6 mm) and tetracycline (10 mm).	[72]

Scientific Name	Part of plant	Family Name	Origin of plant	Country of study	Bacteria	Result	Ref.
Azardirachtaindica	Foliage	meliaceae	India	India	Clinical isolates of Streptococcus mutans	Antibacterial property of aqueous extract of this plant was proved with the inhibition zone diameter of 11.6 mm. However, the inhibition zone diameter of following antibiotic controls was as follows: ampicillin (10 mm), gentamicin (12.3 mm), carbenicillin (13 mm), nalidixic acid (11 mm), nitrofurantoin (10.6 mm) and tetracycline (10 mm).	[72]
Acacia nilotica	Foliage	Fabaceae	India	India	Clinical isolates of Streptococcus mutans	Antibacterial property of aqueous extract of this plant was proved with the inhibition zone diameter of 19.3 mm. However, the inhibition zone diameter of following antibiotic controls was as follows: ampicillin (10 mm), gentamicin (12.3 mm), carbenicillin (13 mm), nalidixic acid (11 mm), nitrofurantoin (10.6 mm) and tetracycline (10 mm).	[72]
Ocimumbasilicum	Foliage	Lamiaceae	India	India	Clinical isolates of Streptococcus mutans	Antibacterial property of aqueous extract of this plant was proved with the inhibition zone diameter of 14 mm. However, the inhibition zone diameter of following antibiotic controls was as follows: ampicillin (10 mm), gentamicin (12.3 mm), carbenicillin (13 mm), nalidixic acid (11 mm), nitrofurantoin (10.6 mm) and tetracycline (10 mm).	[72]
Emblica officinalis	Foliage	Euphorbiaceae	India	India	Clinical isolates of Streptococcus mutans	Antibacterial property of aqueous extract of this plant was proved with the inhibition zone diameter of 29.6 mm. However, the inhibition zone diameter of following antibiotic controls was as follows: ampicillin (10 mm), gentamicin (12.3 mm), carbenicillin (13 mm), nalidixic acid (11 mm), nitrofurantoin (10.6 mm) and tetracycline (10 mm).	[72]
Hemidesmusindicus	Foliage	Asclepiadaceae	India	India	Clinical isolates of Streptococcus mutans	Antibacterial property of aqueous extract of this plant was proved with the inhibition zone diameter of 10.3 mm. However, the inhibition zone diameter of following antibiotic controls was as follows: ampicillin (10 mm), and tetracycline (10 mm).	[72]
Terminaliabellirica	Foliage	Combretaceae	India	India	Clinical isolates of Streptococcus mutans	Antibacterial property of aqueous extract of this plant was proved with the inhibition zone diameter of 33.6 mm. However, the inhibition zone diameter of following antibiotic controls was as follows: ampicillin (10 mm), gentamicin (12.3 mm), carbenicillin (13 mm), nalidixic acid (11 mm), nitrofurantoin (10.6 mm) and tetracycline (10 mm).	[72]

Scientific Name	Part of plant	Family Name	Origin of plant	Country of study	Bacteria	Result	Ref.
Syzygiumcuminii	Foliage	Myrtaceae	India	India	Clinical isolates of Streptococcus mutans	Antibacterial property of aqueous extract of this plant was proved with the inhibition zone diameter of 26.3 mm. However, the inhibition zone diameter of following antibiotic controls was as follows: ampicillin (10 mm), gentamicin (12.3 mm), carbenicillin (13 mm), nalidixic acid (11 mm), nitrofurantoin (10.6 mm) and tetracycline (10 mm).	[72]
Baeckeafrutescens	Leaf		Indone sia	South Korea	Streptococcus mutans ATCC 25175	Methanol extract of this plant showed strong antibacterial property at a concentration of 20 mg / ml with zone of inhibition of 14 mm.	[73]
Glycyrrhizaglabra	Root	Fabaceae	Indone sia	South Korea	Streptococcus mutans ATCC 25175	Methanol extract of this plant showed strong antibacterial property at a concentration of 20 mg / ml with zone of inhibition of 20 mm. Rapid bactericidal effect of this extract also be observed at a concentration of 50 µg / ml within 2 minutes.	[73]
Kaempferiapandurata	Root	Zingiberaceae	Indone sia	South Korea	Streptococcus mutans ATCC 25175	Methanol extract of this plant showed strong antibacterial property at a concentration of 20 mg / ml with zone of inhibition of 14 mm. Rapid bactericidal effect of this extract must also be observed at a concentration of 50 µg / ml within 2 minutes.	[73]
Physalisangulata	Flower	Solanaceae	Indone sia	South Korea	Streptococcus mutans ATCC 25175	Methanol extract of the plant at a concentration of 20 mg / ml with a diameter of 21 mm zone of inhibition showed strong antibacterial properties. Rapid bactericidal effect of this extract at a concentration of 50 μ g / ml within 2 minutes must also be observed.	[73]
Quercusinfectoria	Gall	Fagaceae	Indone sia	South Korea	Streptococcus mutans ATCC 25175	Methanol extract of this plant showed strong antibacterial property at a concentration of 20 mg / ml with zone of inhibition of 22 mm.	[73]
SchinusterebinthifoliusR addi	Barks	Anacardiaceae	Brazil	Brazil	Streptococcusmutans (ATCC25175)	Antibacterial property of ethanol extract of this plant was proved at a concentration of 100 mg / ml with an inhibition zone of 20 mm. However, inhibition zone diameter of 20 mm was also observed for antibiotic control of 0.12% chlorhexidine.	[74]
Psidiumguajava L.	leaves	Myrtaceae	Brazil	Brazil	Streptococcusmutans (ATCC25175)	Antibacterial property of ethanol extract of this plant was proved at a concentration of 100 mg / ml with an inhibition zone of 19 mm. However, inhibition zone diameter of 20 mm was also observed for antibiotic control of 0.12% chlorhexidine.	[74]
Punicagranatum L.	leaves	Punicaceae	Brazil	Brazil	Streptococcusmutans (ATCC25175)	Antibacterial property of ethanol extract of this plant was proved at a concentration of 100 mg / ml with an inhibition zone of 18 mm. However, inhibition zone diameter of 20 mm was also observed for antibiotic control of 0.12% chlorhexidine.	[74]

Scientific Name	Part of plant	Family Name	Origin of plant	Country of study	Bacteria	Result	Ref.
Annonasenegalensis	Bark	Annonaceae	South Africa	South Africa	Streptococcus mutans (ATCC 25175)	Antibacterial property of ethanol extract of this plant was proved with an inhibition zone of 4.5 mm and MIC and MBC equivalent to 12.5 mg / ml. This is despite the fact that MIC and MBC of chlorhexidine, as the antibiotic control, was equal to 1.6 mg / ml.	[75]
Englerophytummagalis montanum	Bark	Sapotaceae	South Africa	South Africa	Streptococcus mutans (ATCC 25175)	Antibacterial property of ethanol extract of this plant was proved with an inhibition zone of 3.5 mm and MIC (12.5 mg / ml) and MBC (6.3 mg / ml). This is despite the fact that MIC and MBC of chlorhexidine, as the antibiotic control, was equal to 1.6 mg / ml.	[75]
Dicerocarymsenecioides	Roots	Pedaliaceae	South Africa	South Africa	Streptococcus mutans (ATCC 25175)	Antibacterial property of ethanol extract of this plant was proved with an inhibition zone of 3.5 mm and MIC (12.5 mg / ml) and MBC (25 mg / ml). This is despite the fact that MIC and MBC of chlorhexidine, as the antibiotic control, was equal to 1.6 mg / ml.	[75]
Eucleadivinorum	Bark, Leaves	Ebenaceae	South Africa	South Africa	Streptococcus mutans (ATCC 25175)	Antibacterial property of ethanol extract of this plant was proved with an inhibition zone of 6 mm and MIC (25 mg / ml) and MBC (25 mg / ml). This is despite the fact that MIC and MBC of chlorhexidine, as the antibiotic control, was equal to 1.6 mg / ml.	[75]
Eucleanatalensis	Leave s	Ebenaceae	South Africa	South Africa	Streptococcus mutans (ATCC 25175)	Antibacterial property of ethanol extract of this plant was proved with an inhibition zone of 7.6 mm and MIC (6.3 mg / ml) and MBC (6.3 mg / ml). This is despite the fact that MIC and MBC of chlorhexidine, as the antibiotic control, was equal to 1.6 mg / ml.	[75]
Parinaricuratellifolia	Bark	Chrysobalanceae	South Africa	South Africa	Streptococcus mutans (ATCC 25175)	Antibacterial property of ethanol extract of this plant was proved with an inhibition zone of 4.2 mm and MIC (6.3 mg / ml) and MBC (6.3 mg / ml). This is despite the fact that MIC and MBC of chlorhexidine, as the antibiotic control, was equal to 1.6 mg / ml.	[75]

DISCUSSION

Although there are various ways to control infections such as chemotherapeutics which are able to eliminate microbes selectively, but most of microbes get resistant to antibiotics very soon and also they mostly have side effects. A lot of studies on medicinal plant extracts not only have shown antibacterial activities but also they have various other effects [76-85]. Medicinal herbs since ancient times have been a source of traditional medicine and the treatment of diseases. These herbs contain bioactive substances and antioxidant activities [86-89]. Most of them belong to the same therapeutic effects of active pharmaceutical ingredients in them [86-93]. Bioactive compounds from medicinal plants have therapeutic effects on various diseases [94-101].

Today, the incidence of infectious diseases, particularly bacterial diseases have expanded. These batteries induce infections by various mechanisms [102-104]. These diseases have been exposed to drug resistance. A good strategy to address antibiotic resistance is the source of medicinal plants [105]. The extracts of some medicinal plants including Baeckea frutescens, Quercus infectoria Glycyrrhiza glabra, Physalis angulata and Kaempferia pandurata have shown potent antibacterial activities against Streptococcus mutans. P. angulate, G. glabra and K. pandurata, in particular showed fast bactericidal impact on S. mutans at 50 µg/ml of extract, in 2 min [73].

CONCLUSION

The mechanism actions of these plants are not known, however, phenolic compound have shown to have antibacterial activities. Most of the plants presented in this study have phenolic compounds and may have antibacterial activity by these compounds. There are a lot of other plants which have these or similar compounds [106-112]. Hence they also may have antibacterial activities.

REFERENCES:

- Ambrosio SR, Furtado NA, de Oliveira DC, da Costa FB, Martins CH, de Carvalho TC. Antimicrobial activity of kauranediterpenes against oral pathogens. Z Naturforsch C. 2008;63(5-6):326-30.
- [2] Shemesh M, Tam A, Feldman M, Steinberg D. Differential expression profiles of Streptococcus mutans ftf, gtf and vicR genes in the presence of dietary carbohydrates at early and late exponential growth phases. Carbohydr Res2006 4; 341(12):2090-7.
- [3] Maghareabed A, Bateni E, Rabiei A, Pour Moradi B. The Review on the Effect of Using Mouthwash in Keeping Mouth and Teeth Healthy. Isfahan Dentistry Faculty Journal.2012;7(5).
- [4] Kuhnert WL, Quivey Jr RG Jr.Genetic and Biochemical Characterization of the F-TPaseoperon from Streptococcus sanguis 10904. J Bacteriol. 2003;185(5):1525-33.
- [5] Takahashi N, Nyvad B. The role of Bacteria in the Caries Process Ecological Perspectives. J Dent Res2011;90(3):294-303.
- [6] Thorild I, Lindau-Janson B, Twetman S. Prevalence of salivary Streptococcus mutans in mothers and in their preschool children. Int J Paediatr Dent 2002; 12(1): 2-7.
- [7] Kreth J, Merritt J, Qi F. Bacterial and host interactions of oral streptococci. DNA cell biology 2009;
- [8] Moori JJ, Nan HJ, steal JG. Prevention of oral disease. 1th ed. Tehran: Hampa publications; 2009. p. 92-5.
- [9] Kuramitsu HK. Virulence factors of mutans streptococci: role of molecular genetics. Critical Review of Oral Biological Medicine 1993; 4: 159-176.
- [10] Rezvanirad A, Mardani M, Shirzad H, Ahmadzadeh SM, Asgary S, Naimi A, Mahmoudi GHA. Curcuma longa: A review of therapeutic effects in traditional and modern medical references. J Chem Pharmac Sci. 2016; 9 (4): 3438-3448.
- [11] Mahmoudi GA, Mahmoodnia L, Mirhosseini M. Medicinal plants with anti-poisoning toxicity of carbon tetrachloride: An overview of the most important medicinal plants native to Iran with anti- carbon tetrachloride toxicity. J Global Pharma Technol. 2016; 8 (11): 17-20.
- [12] Bahmani M, Karamati SA, Hassanzadazar H, Forouzan SH, Rafieian-Kopaei M, Kazemi-Ghoshchi B, Asadzadeh J, Kheiri AGh, Ehsan Bahmani E. Ethnobotanic study of medicinal plants in Urmia city: identification and traditional using of antiparasites plants. Asian Pac J Trop Dis 2014; 4(Suppl 2): 906-910.
- [13] Nasri H, Shirzad H, Baradaran A. Rafieian-kopaei M. Antioxidant plants and diabetes mellitus. J Res Med Sci 2015; 20:491-50
- [14] Mahmoudi GA, Mahmoodnia L, Mirhosseini M. A review on the most important medicinal herbs native to Iran with antiacetaminophen toxicity. J Global Pharma Technol. 2016; 8 (11): 12-16.
- [15] Mahmoudi GA, Almasi V, Lorzadeh N, Khansari A. The reasons for using and not using alternative medicine in Khorramabad women, west of Iran. J Pakistan Med Assoc. 2015; 65(6): 623-625.
- [16] Parsaei P, Karimi M, Asadi SY, Rafieian-Kopaei M. Bioactive components and preventive effect of green tea (Camellia sinensis) extract on postlaparotomy intra-abdominal adhesion in rats. Int J Surg. 2013; http://dx.doi.org/10.1016/j.ijsu.2013.08.014.
- [17] Rabiei Z, Rafieian-Kopaei M, Heidarian E, Saghaei E, Mokhtari S.Effects of Zizyphus jujube extract on memory and learning impairment induced by bilateral electric lesions of the nucleus Basalis of Meynert in rat. Neurochem Res. 2014 Feb;39(2):353-60.
- [18] Shirzad H, Shahrani M, Rafieian-Kopaei M. Comparison of morphine and tramadol effects on phagocytic activity of mice peritoneal phagocytes in vivo. Int Immunopharmacol. 2009 Jul;9(7-8):968-70. Epub 2009 Apr 8. PMID: 19361579.
- [19] Rafieian-Kopaei M, Asgary S, Adelnia A, Setorki M, Khazaei M, Kazemi S, Shamsi F. The effects of cornelian cherry on atherosclerosis and atherogenic factors in hypercholesterolemic rabbits. J Med Plants Res. 2011; 5(13): 2670-2676.
- [20] Bahmani M, Shirzad H, Rafieian S, Rafieian-Kopaei M. Silybum marianum: Beyond Hepatoprotection. J Evidence-Based Complement Alternative Med 2015; 20(4): 292-301.
- [21] Bahmani, M., Rafieian-Kopaei, M. Medicinal plants and secondary metabolites for leech control. Asian Pac J Trop Dis. 2014; 4(4): 315-316.
- [22] Bahmani M, Sarrafchi A, Shirzad H, Rafieian-Kopaei M. Autism: Pathophysiology and promising herbal remedies. Current Pharmaceutical Design 2016; 22(3): 277-285

- [23] Sewell RDE, Rafieian-Kopaei M. The history and ups and downs of herbal medicine usage. J Herbmed Pharmacol. 2014; 3(1): 1-3.
- [24] Rezapour S, Bahmani M, Afsordeh O, Rafieian R, Sheikhian A. Herbal medicines: a new hope for autism therapy. J Herbmed Pharmacol. 2016;5(3):89-91.
- [25] Asadi-Samani M, Kooti W, Aslani E, Shirzad H: A systematic review of Iran's medicinal plants with anticancer effects. Journal of Evidence-Based Complementary & Alternative Medicine. 2016; 21(2): 143-153.
- [26] Ahmadipour S, Ahmadipour S, Mohsenzadeh A, Asadi-Samani M. The importance of some native medicinal plants of Iran effective on gastrointestinal disorders in children: A review. Der Pharmacia Lettre. 2016; 8(1):61-6.
- [27] Ebrahimie M, Bahmani M, Shirzad H, Rafieian-Kopaei M, Saki K. A Review Study on the Effect of Iranian Herbal Medicines on Opioid Withdrawal Syndrome. J Evid Based Complementary Altern Med. 2015 Oct;20(4):302-9. doi: 10.1177/2156587215577896. Epub 2015 Mar 26
- [28] Ganji-Arjenaki M, Rafieian-Kopaei M. Probiotics are a good choice in remission of inflammatory bowel diseases: A Meta Analysis and systematic review. Journal of Cellular Physiology. 2017 Mar 15. PubMed PMID: 28294322.
- [29] Ghaed F, Rafieian-Kopaei M, Nematbakhsh M, Baradaran A, Nasri H. Ameliorative effects of metformin on renal histologic and biochemical alterations of gentamicin-induced renal toxicity in Wistar rats. J Res Med Sci. 2012; 17 (7): 621-625.
- [30] Gharipour M, Ramezani MA, Sadeghi M, Khosravi A, Masjedi M, Khosravi-Boroujeni H.et al. Sex based levels of C reactive protein and white blood cell count in subjects with metabolic syndrome: Isfahan Healthy Heart Program. J Res Med Sci 2013;18:467-72.
- [31] Heidarian E, Rafieian-Kopaei M. Protective effect of artichoke (Cynara scolymus) leaf extract against lead toxicity in rat. Pharm Biol. 2013 Sep;51(9):1104-9. doi: 10.3109/13880209.2013.777931. Epub 2013 Jun 7.
- [32] Bahmani M, Shirzad H, Mirhosseini M, Mesripour A, Rafieian-Kopaei M. A Review on Ethnobotanical and Therapeutic Uses of Fenugreek (Trigonella foenum-graceum L). J Evid Based Complementary Altern Med. 2015 Apr 27. pii: 2156587215583405. [Epub ahead of print].
- [33] Bahmani M, Shirzad H, Rafieian S, Rafieian-Kopaei M. Silybum marianum: Beyond Hepatoprotection. J Evid Based Complementary Altern Med. 2015, 20(4) 292-301.
- [34] Bahmani M, Shirzad HA, Majlesi M, Shahinfard N, Rafieian-Kopaei M. A review study on analgesic applications of Iranian medicinal plants. Asian Pac J Trop Med 2014; 7(Suppl 1): 43-53.
- [35] Kafash-Farkhad N, Asadi-Samani M, Rafieian-Kopaei M. A review on phytochemistry and pharmacological effects of Prangos ferulacea (L.) Lindl. Life Sci J. 2013; 10(8s):360-367
- [36] Mohsenzadeh A, Ahmadipour S, Ahmadipour S, Asadi-Samani M. A review of the most important medicinal plants effective on cough in children and adults. Der Pharmacia Lettre. 2016;8(1):90-6.
- [37] Khosravi-Boroujeni H, Mohammadifard N, Sarrafzadegan N, Sajjadi F, Maghroun M, Khosravi A, Alikhasi H, Rafieian M, Azadbakht L. Potato consumption and cardiovascular disease risk factors among Iranian population. Int J Food Sci Nutr. 2012; 63(8):913-20.
- [38] Bahmani M, Zargaran A, Rafieian-Kopaei M, Saki M. Ethnobotanical study of medicinal plants used in the management of diabetes mellitus in the Urmia, Northwest Iran. Asian Pac J Trop Med 2014; 7(Suppl 1): 348-354.
- [39] Bahmani M, Zargaran A, Rafieian-Kopaei M. Identification of medicinal plants of Urmia for treatment of gastrointestinal disorders. Rev Bras Farmacogn 24(2014): 468-480
- [40] Bahmani M, Rafieian-Kopaei M. Medicinal plants and secondary metabolites for leech control. Asian Pacific J Trop Dis 2014; 4(4): 315-316.
- [41] Khosravi-Boroujeni H, Sarrafzadegan N, Mohammadifard N, Sajjadi F, Maghroun M, Asgari S, Rafieian-Kopaei M, Azadbakht L. White rice consumption and CVD risk factors among Iranian population. J Health Popul Nutr. 2013 Jun;31(2):252-61.
- [42] Baharvand-Ahmadi B, Bahmani M, Tajeddini P, Naghdi N, Rafieian-Kopaei M. An ethno-medicinal study of medicinal plants used for the treatment of diabetes. J Nephropathol. 2016; 5(1):44-50.
- [43] Baharvand-Ahmadi, B., Bahmani, M., Naghdi, N., Saki, K., Baharvand-Ahmadi, S., Rafieian-Kopaei, M. Review on

phytochemistry, therapeutic and pharmacological effects of myrtus (Myrtus communis). Der Pharmacia Lettre 2015; 7(11): 160-165.

- [44] Bahmani M, Rafieian M, Baradaran A, Rafieian S, Rafieian-kopaei M. Nephrotoxicity and hepatotoxicity evaluation of Crocus sativus stigmas in neonates of nursing mice. J Nephropathol. 2014; 3(2): 81-85.
- [45] Bahmani M, Rafieian-Kopaei M, Hassanzadazar H, Saki K, Karamati SA, Delfan B. A review on most important herbal and synthetic antihelmintic drugs. Asian Pac J Trop Med 2014; 7(Suppl 1): 29-33.
- [46] Bahmani M, Rafieian-Kopaei M, Jeloudari M, Eftekhari Z, Delfan B, Zargaran A, Forouzan SH. A review of the health effects and uses of drugs of plant licorice (Glycyrrhiza glabra L.) in Iran. Asian Pac J Trop Dis 2014; 4(Suppl 2): 847-849.
- [47] Shirzad H, Taji F, Rafieian-Kopaei M. Correlation between antioxidant activity of garlic extracts and WEHI-164 fibrosarcoma tumor growth in BALB/c mice. J Med Food. 2011 Sep; 14(9):969-74.
- [48] Rafieian-Kopaie M, Nasri H, Alizadeh F, Ataebi B, Baradaran A. Immunoglobulin A nephropathy and Malaria falciparum infection; a rare association. Iranian J Pub Health 2013. 42(5):529-533.
- [49] Shokri H, Sharifzadeh A. Zataria multiflora Boiss.: A review study on chemical composition, antifungal and anti-mycotoxin activities, and ultrastructural changes. J Herbmed Pharmacol. 2017;6(1):1-9.
- [50] Ihekwereme CP, Agbata CA, Chukwueze KO, Agu SC. In vivo evaluation of antiplasmodial activity of hydroethanolic stem extract of Baphia pubescens in Plasmodium berghei infected albino mice. J HerbMed Pharmacol. 2016;5(4):149-152.
- [51] Rafiean-Kopaei M, Baradaran A, Maghsoudi AR, Ghobadi Sh, Nasri H. Helicobacter pylori infection and serum homocysteine in hemodialysis patient. Life Sci J. 2012; 9(4): 3696-3702.
- [52] Asadi-Samani M, Bahmani M, Rafieian-Kopaei M. The chemical composition, botanical characteristic and biological activities of Borago officinalis: a review. Asian Pac J Trop Med 2014; 7(Suppl 1): 22-28.
- [53] Bahmani M, Saki K, Rafieian-Kopaei M, Karamati SA, Eftekhari Z, Jelodari M. The most common herbal medicines affecting Sarcomastigophora branches: a review study. Asian Pac J Trop Med 2014; 7(Suppl 1): 14-21.
- [54] Delfan B, Kazemeini H, Bahmani M. Identifying effective medicinal plants for cold in Lorestan province, West of Iran. J Evid Based Complementary Altern Med. 2015 Jul;20(3):173-9. doi: 10.1177/2156587214568458. Epub 2015 Jan 22.
- [55] Akhlaghi M, Shabanian Gh, Rafieian-Koupaei M, Parvin N, Saadat M, Akhlaghi M. Citrus aurantium Blossom and Preoperative Anxiety. Revista Brasileira de Anestesiologia 2011; 61(6):702-712.
- [56] Amirmohammadi M, Khajoenia S, Bahmani M, Rafieian-Kopaei M, Eftekhari Z, Qorbani M. In vivo evaluation of antiparasitic effects of Artemisia abrotanum and Salvia officinalis extracts on Syphacia obvelata, Aspiculoris tetrapetra and Hymenolepis nana parasites. Asian Pac J Trop Dis. 2014; 4(Suppl 1): S250-S254.
- [57] Bagheri N, Rahimian Gh, Salimzadeh L, Azadegan F, Rafieian-Kopaei M, Taghikhani A, Shirzad H .Association of the Virulence factors of Helicobacter pylori and Gastric Mucosal Interleukin-17/23 mRNA Expression in dyspeptic patients. EXCLI J. 2013; 12:5-14.
- [58] Bagheri N, Taghikhani A, Rahimian G, Salimzadeh L, Azadegan Dehkordi F, Zandi F, Chaleshtori MH, Rafieian-Kopaei M, Shirzad H. Association between virulence factors of helicobacter pylori and gastric mucosal interleukin-18 mRNA expression in dyspeptic patients. Microb Pathog. 2013 Dec;65:7-13. doi: 10.1016/j.micpath.2013.08.005. Epub 2013 Sep 10.
- [59] Jebashree HS, Kingsley SJ, Sathish ES, Devapriya D. Antimicrobial activity of few medicinal plants against clinically isolated human cariogenic pathogens—An in vitro study. ISRN dentistry. 2011;--.
- [60] Song J-H, Kim S-K, Chang K-W, Han S-K, Yi H-K, Jeon J-G. In vitro inhibitory effects of Polygonum cuspidatum on bacterial viability and virulence factors of Streptococcus mutans and Streptococcus sobrinus. Archives of Oral Biology. 2006;51(12):1131-40.
- [61] Chaudhry NMA, Tariq P. Anti-microbial activity of Cinnamomum cassia against diverse microbial flora with its nutritional and medicinal impacts. Pakistan Journal of Botany. 2006;38(1):169.
- [62] Yatsuda R, Rosalen P, Cury J, Murata R, Rehder V, Melo L, et al. Effects of Mikania genus plants on growth and cell adherence of

mutans streptococci. Journal of Ethnopharmacology. 2005;97(2):183-9.

- [63] Chaiya A, Saraya S, Chuakul W, Temsiririrkkul R. Screening for dental caries: Preventive activities of medicinal plants against Streptococcus mutans. Mahidol University Journal of Pharmaceutical Sciences. 2013;40(1):9-17.
- [64] Tsai T-H, Tsai T-H, Chien Y-C, Lee C-W, Tsai P-J. In vitro antimicrobial activities against cariogenic streptococci and their antioxidant capacities: A comparative study of green tea versus different herbs. Food Chemistry. 2008;110 (4):859-64.
- [65] Rosas-Piñón Y, Mejía A, Díaz-Ruiz G, Aguilar MI, Sánchez-Nieto S, Rivero-Cruz JF. Ethnobotanical survey and antibacterial activity of plants used in the Altiplane region of Mexico for the treatment of oral cavity infections. Journal of ethnopharmacology. 2012; 141(3):860-5.
- [66] Ocheng F, Bwanga F, Joloba M, Borg-Karlson A-K, Gustafsson A, Obua C. Antibacterial activities of extracts from Ugandan medicinal plants used for oral care. Journal of ethnopharmacology. 2014; 155(1):852-5.
- [67] Wang J-K, Chung J-Y, Baek N-I, Park J-H. Isopanduratin A from Kaempferia pandurata as an active antibacterial agent against cariogenic Streptococcus mutans. International journal of antimicrobial agents. 2004;23(4):377-81.
- [68] Almeida LSBd, Murata RM, Yatsuda R, Dos Santos M, Nagem TJ, Alencar SMd, et al. Antimicrobial activity of Rheedia brasiliensis and 7-epiclusianone against Streptococcus mutans. Phytomedicine. 2008;15(10):886-91.
- [69] Debnath M. Clonal propagation and antimicrobial activity of an endemic medicinal plant Stevia rebaudiana. Journal of medicinal plants research. 2007;2(2):045-51.
- [70] Sharma UK, Sharma AK, Pandey AK. Medicinal attributes of major phenylpropanoids present in cinnamon. BMC complementary and alternative medicine. 2016;16(1):1.
- [71] Al-Bayati FA, Sulaiman KD. In vitro antimicrobial activity of Salvadora persica L. extracts against some isolated oral pathogens in Iraq. Turkish Journal of Biology. 2008; 32(1):57-62.
- [72] Pathak A, Sardar A, Kadam V, Rekadwad B, Karuppayil SM. Efficacy of some medicinal plants against human dental pathogens. Indian Journal of Natural Products and Resources. 2012; 3(1):123-7.
- [73] Hwang J-K, Shim J-S, Chung J-Y. Anticariogenic activity of some tropical medicinal plants against Streptococcus mutans. Fitoterapia. 2004;75(6):596-8.
- [74] Vieira DR, Amaral FM, Maciel MC, Nascimento FR, Libério SA, Rodrigues VP. Plant species used in dental diseases: Ethnopharmacology aspects and antimicrobial activity evaluation. Journal of ethnopharmacology. 2014;155(3):1441-9.
- [75] More G, Tshikalange TE, Lall N, Botha F, Meyer JJM. Antimicrobial activity of medicinal plants against oral microorganisms. Journal of Ethnopharmacology. 2008;119(3):473-7.
- [76] Bahmani M, Mirhoseini M, Shirzad H, Sedighi M, Shahinfard N, Rafieian-Kopaei M. A review on promising natural agents effective on hyperlipidemia. J Evid Based Complementary Altern Med. 2015 Jul;20(3):228-38. doi: 10.1177/2156587214568457. Epub 2015 Jan 28
- [77] Bahmani M, Sarrafchi A, Shirzad H, Rafieian-Kopaei M. Autism: Pathophysiology and promising herbal remedies. Curr Pharm Des. 2016; 22(3):277–285. DOI: 10.2174/1381612822666151112151529
- [78] Delfan B, Bahmani M, Rafieian-Kopaei M, Delfan M, Saki K. A review study on ethnobotanical study of medicinal plants used in relief of toothache in Lorestan Province, Iran. Asian Pac J Trop Dis 2014; 4(Suppl 2): 879-884.
- [79] Saki K, Bahmani M, Rafieian-Kopaei M, Hassanzadazar H, Dehghan K, Bahmani F, Asadzadeh J. The most common native medicinal plants used for psychiatric and neurological disorders in Urmia city, northwest of Iran. Asian Pac J Trop Dis 2014; 4(Suppl 2): 895-901.
- [80] Sarrafzadegan N, Khosravi-Boroujeni H, Esmaillzadeh A, Sadeghi M, Rafieian-Kopaei, M., Asgary S. The association between hypertriglyceridemic waist phenotype, menopause, and cardiovascular risk factors. Arch Iran Med. 2013; 16(3):161-166.
- [81] Sedighi M., Rafieian-Kopaei M., Noori-Ahmadabadi M. Kelussia odoratissima Mozaffarian inhibits ileum contractions through voltage dependent and beta adrenergic receptors. Life Sci J. 9(4) 2012: 1033-1038.

- [82] Saki K, Bahmani M, Rafieian-Kopaei M. The effect of most important medicinal plants on two important psychiatric disorders (anxiety and depression)-a review. Asian Pac J Trop Med 2014; 7(Suppl 1): 34-42.
- [83] Salehi A, Jabarzare S, Neurmohamadi M, Kheiri S, Rafieian-Kopaei M. A double blind clinical trial on the efficacy of honey drop in vernal keratoconjunctivitis. Evid Based Complement Alternat Med. 2014;2014:287540. doi: 10.1155/2014/287540. Epub 2014 Feb 24.
- [84] Sharafati R, Sharafati F, Rafieian-kopaei M. Biological characterization of Iranian walnut (Juglans regia) leaves. Turk J Biol. 2011:635-9.
- [85] Shayganni E, Bahmani M, Asgary S, Rafieian-Kopaei M. Inflammaging and cardiovascular disease: management by medicinal plants. Phytomedicine. 2016; 23: 1119–1126.
- [86] Lorigooini Z, Kobarfard F, Ayatollahi SA. Anti-platelet aggregation assay and chemical composition of essential oil from Allium atroviolaceum Boiss growing in Iran. International Journal of Biosciences (IJB). 2014;5(2):151-6.
- [87] Lorigooini Z, Ayatollahi SA, Amidi S, Kobarfard F. Evaluation of anti-platelet aggregation effect of some Allium species. Iranian journal of pharmaceutical research: IJPR. 2015;14(4):1225.
- [88] Fasihzadeh, S, Lorigooini, Z, Jivad, N. Chemical constituents of Allium stipitatum regel (persian shallot) essential oil. Der Pharmacia Lettre. 2016;8 (1):175-180.
- [89] Ghasemi S, Lorigooini Z. A review of significant molecular mechanisms of flavonoids in prevention of prostate cancer. Journal of Chemical and Pharmaceutical Sciences. 2016;9: 3388-3394.
- [90] Asgary S, Kelishadi R, Rafieian-Kopaei M, Najafi S, Najafi M, Sahebkar A. Investigation of the lipid-modifying and antiinflammatory effects of Cornus mas L. supplementation on dyslipidemic children and adolescents. Pediatr Cardiol. 2013 Oct;34(7):1729-35. doi: 10.1007/s00246-013-0693-5. Epub 2013 Apr 27.
- [91] Noori-Ahmad-Abadi M, Mohammad Reza Hojjati, Mehrnoosh Sedighi. Effect of hydro-alcoholic extract of Ziziphus Jujuba on the peripheral blood cells in Balb/c mice. Physiology and Pharmacology. 2013; 17 (2), 224-230.
- [92] Sarrafzadegan N, Khosravi-Boroujeni H, Esmaillzadeh A, Sadeghi M, Rafieian-Kopaei, M., Asgary S. The association between hypertriglyceridemic waist phenotype, menopause, and cardiovascular risk factors. Arch Iran Med. 2013; 16(3):161-166.
- [93] Rafieian-Kopaei M, Baradaran A, Merrikhi A, Nematbakhsh M, Madihi Y, Nasri H. Efficacy of Co-Administration of garlic extract and metformin for prevention of gentamicin-renal toxicity in wistar rats: A biochemical study. International Journal of Preventive Medicine. 2013;4(3):258-64.
- [94] Kooti W, Ghasemiboroon M, Ahangarpoor A, Hardani A, Amirzargar A, Asadi-Samani M, et al. The effect of hydro-alcoholic extract of celery on male rats in fertility control and sex ratio of rat offspring. Journal of Babol University of Medical Sciences. 2014;16(4):43-9.
- [95] Jivad N, Bahmani M, Asadi-Samani M. A review of the most important medicinal plants effective on wound healing on ethnobotany evidence of Iran. Der Pharmacia Lettre. 2016;8(2):353-7.
- [96] Parsaei P, Bahmani M, Karimi M, Naghdi N, Asadi-Samani M, Rafieian-Kopaei M. A review of analgesic medicinal plants in Iran. Der Pharmacia Lettre. 2016;8(2):43-51.
- [97] Kooti W, Ghasemiboroon M, Asadi-Samani M, Ahangarpoor A, Abadi MNA, Afrisham R, et al. The effects of hydro-alcoholic

extract of celery on lipid profile of rats fed a high fat diet. Advances in Environmental Biology. 2014;8(4):325-30.

- [98] Jivad N, Asadi-Samani M, Moradi MT. The most important medicinal plants effective on migraine: A review of ethnobotanical studies in Iran. Der Pharma Chemica. 2016;8(2):462-6.
- [99] Rafieian-Kopaei M, Baradaran A, Nasri H. Significance of extracapillary proliferation in IgA-nephropathy patients with regard to clinical and histopathological variable. Hippokratia 2013, 17, 3: 258-261.
- [100] Sedighi M., Rafieian-Kopaei M., Noori-Ahmadabadi M. Kelussia odoratissima Mozaffarian inhibits ileum contractions through voltage dependent and beta adrenergic receptors. Life Sci J. 9(4) 2012: 1033-1038.
- [101] Seddighi M, Bahmani M, Asgary S, Beyranvand F, Rafieian-Kopaei M. A review of plant-based compounds and medicinal plants effective on atherosclerosis. J Res Med Sci. 2017; 22:30.
- [102] Bagheri N, Azadegan-Dehkordi F, Rahimian G, Hashemzadeh-Chaleshtori M, Rafieian-Kopaei M, Kheiri S, et al. Altered Th17 Cytokine Expression in Helicobacter pylori Patients with TLR4 (D299G) Polymorphism. Immunol Invest. 2016:1-11.
- [103] Bagheri N, Azadegan-Dehkordi F, Shirzad M, Zamanzad B, Rahimian G, Taghikhani A, et al. Mucosal interleukin-21 mRNA expression level is high in patients with Helicobacter pylori and is associated with the severity of gastritis. Centr Eur Immunol. 2015;40:61-7.
- [104] Razavi A, Bagheri N, Azadegan-Dehkordi F, Shirzad M, Rahimian G, Rafieian-Kopaei M, et al. Comparative Immune Response in Children and Adults with H. pylori Infection. Journal of Immunology Research. 2015;2015:315957.
- [105] Zomorodian K, Moein M, Lori ZG, Ghasemi Y, Rahimi MJ, Bandegani A, Pakshir K, Bazargani A, Mirzamohammadi S, Abbasi N. Chemical composition and antimicrobial activities of the essential oil from Myrtus communis leaves. Journal of Essential Oil Bearing Plants. 2013;16(1):76-84.
- [106]Esehaghbeygi, A., Hoseinzadeh, B., Masoumi, A.A. Effects of moisture content and urea fertilizer on bending and shearing properties of canola stem. Applied Engineering in Agriculture. 2009;25(6):947-951.
- [107] Hoseinzadeh, B., Esehaghbeygi, A., Raghami, N. Silique picking force for canola. International Journal of Agriculture and Biology. 2010;12 (4): 632-634.
- [108] Ghazavi, M.A., Mohammadi, A., Hosseinzadeh Samani.B. Comparison of some of mechanical properties of Shahrud 12 and Mama'e almonds under pressure loading. Journal of Food, Agriculture and Environment. 2011;9(2):257-262.
- [109] Sarrafchi A, Bahmani M, Shirzad H, Rafieian-Kopaei M. Oxidative stress and Parkinson's disease: New hopes in treatment with herbal antioxidants. Curr Pharm Des. 2016; 22(2): 238 – 246. DOI: 10.2174/1381612822666151112151653
- [110] Rabie Z, Gholami M, Rafieian-Kopaei M. Antidepressant effects of Mentha pulegium in mice. Bangladesh J Pharmacol. 2016; 11(3): 711-715 doi:http://dx.doi.org/10.3329/bjp.v11i3.27318.
- [111] Sharafati-Chaleshtori R, Shirzad H, Rafieian-Kopaei M, Soltani A. Melatonin and human mitochondrial diseases. J Res Med Sci 2016;21:138.
- [112] Rouhi-Boroujeni H, Heidarian E, Rouhi-Boroujeni H, Deris F, Rafieian-Kopaei M. Medicinal Plants with multiple effects on cardiovascular diseases: a systematic review. Curr Pharm Des. 2017; 23(7): 999 – 1015. DOI: 10.2174/1381612822666161021160524.