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# A review on biosynthesis, characterization and Antimicrobial effect of silver nanoparticles of *Moringa olifera* (MO-AgNPs)

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# Abstract:

Some common plants in India rich in a wide variety of secondary metabolites, viz. tannins, terpenoids, alkaloids, flavonoids, quinines, phenols.  $\beta$ -sitosterol, coffeoylquinic acid, quercetin and kaemperol. The traditional medicinal plant *Moringa oleifera* is an integral part of the Indian diet and has notable beneficial effects in its leaves, stems, flowers, roots, bark and seeds. It has reported properties like antimicrobial, anti-inflammatory, ant-diabetic, anti-oxidative, anti-tumorogenic amongst many other properties. The major challenge the world is facing today is the mode of treatment of pathogenic bacteria which have become resistant to the existing antibiotics. Green synthesis of metal nanoparticles has become an important branch of nanotechnology and there is an increasing commercial demand for nanoparticles. Nanoparticles have high penetrating ability than the antibiotics. Among all the nanoparticles (NPs), silver nanoparticles (AgNPs) are one of the promising nano product widely used in the field of nanomedicine. The present review describes *Moringa olifera* silver nanoparticles (MO-AgNPs) can be used as new novel source of antimicrobics to combat multiple drug resistant tough microorganisms.

Key words: Moringa oleifera, Silver nanoparticles, Characterization, Antimicrobial properties.

# 1. Introduction:

In India around 8000 species of plants are reported which have medicinal properties and designated as medicinal plants (Agarwal and Gayathri, 2017). Medicinal plants have provided to be the best for the treatment of disease (Silva and Junior, 2010). Plant extracts has been demonstrated to be high in antioxidant activity and is effective in the prevention of atherosclerosis, coronary heart disease, cancer and a number of other diseases (Fuhrman, 2005, Sumner, 2005). There are various reports about the presence of secondary metabolites like tannins, alkaloids, glycosides, flavonoids and phenolic compounds as antioxidant factors in different plant materials. Again, overproduction of free radicals in certain conditions can cause an imbalance, leading to oxidative damage to large biomolecules such as lipids, DNA, and proteins (Liu, 2002) and thus leads to a range of chronic diseases, such as cardiovascular disease, neuronal disease, cataracts, and several forms of cancer (Halliwell, 1997). Moringa oleifera is a perennial softwood tree with timber of low quality, but which for centuries has been advocated for traditional medicinal industrial uses (Fahev. 2005). Nanotechnology is now creating a growing sense of excitement in life sciences especially biomedical devices and biotechnology (Prabhu, 2010). Nanoparticles (NPs) exhibit completely new or improved properties based on specific characteristics such as size, distribution and morphology. Metal nanoparticles have received significant attention in recent years owing to their unique properties and practical applications (Ahmad et al., 2003; Shahverdi, 2007; Bhusnure et al., 2017). Green nanotechnology is an area with significant focus at present on the important facilitating objective of the manufacture nanotechnology-based products that are eco-friendly and safer for all beings, with sustainable commercial viability. The green synthesis of metal nanoparticles receives great attention due to their unusual optical, chemical, photochemical, and electronic properties (Mohanpuria et al., 2008).

The silver nanoparticles have various applications in several ways historical; silver has been known to have a disinfecting effect and has been found in applications ranging from traditional medicines to culinary items. It has been reported that silver nanoparticles (AgNPs) are non-toxic to humans and most effective against bacteria, virus and other micro-organisms at low concentrations and without any side effects (Jeong et al., 2005). Plant extracts have shown large prospects in silver nanoparticle (AgNP) synthesis (Ghosh et al., 2014). Silver nanoparticles synthesized from plant extracts have important applications in biology such as antibacterial agents and DNA sequencing. Antimicrobial property of silver nanoparticles against Staphyloccocus aureus, Pseudomonas aeruginosa and Escherichia coli has been investigated by Rai et al. (2009). AgNPs have been studied because of their strong optical absorption in the visible region caused by the collective excitation of free-electron gas (Mohamed, 2000). AgNPs have a wide area of interest, as they have a large number of applications, such as in nonlinear optics, spectrally selective coating for solar energy absorption, biolabeling, intercalation materials for electrical batteries as optical receptors, catalyst in chemical reactions, and as antibacterial capacities (Sathishkumar, 2009). AgNPs synthesized from Moringa oliefera have important applications in biology such as antibacterial agents. It was also shown that the antibacterial activity of AgNPs was size dependent. AgNPs mainly in the range of 1 -10 nm attach to the surface of cell membrane of the microorganism and drastically disturb its proper function like respiration and permeability (Morones, 2005).

# 2. MEDICINAL PROPERTIES OF MORINGA OLIFERA:

In the last few years various studies has been done for its antimicrobial activity from the extract made using chloroform, ethanol. The ethanolic extract of M. oleifera leaves has been demonstrated to exhibit anthelmintic activity against Indian earthworm (Rastogi et al., 2009), antifungal activity against dermatophytes (Chuang et al., 2007), antifertility (Prakash, 1998; Shukla et al., 1981) and hypoglycemic potential (Jaiswal et al., 2009). Almost all parts of the plant have been utilized in the traditional system of medicine. The plant leaves have been reported for its antitumor, cardioprotective, hypotensive, wound and eye healing properties (Rathi et al., 2006). A study on evaluation of M. oleifera leaves extract on ovariectomy induced bone loss in rats records that the ethanolic extract of M. oleifera leaves possess osteoprotective effect comparable with estradiol (Burali et al., 2010) and has been reported to reduce cyclophosphamide induced immunodepression by stimulating cellular and humoral immunity in mice (Gupta et al., 2010; Siddarth and Gupta, 2007). The aqueous extract of M oleifera leaves have been demonstrated to exhibit protective effect on ulcerated gastric tissue induced by aspirin, cerebral nodular lesion and cold stress in rats (Patel et al., 2008), wound healing property in rats (Makkar and Becker, 1996) significant hypoglycemic and antidiabetic potential (Jaiswal et al., 2009) and the regulatory control on thyroid hormone status in adult Swiss rats (Rathi et al., 2006).

Many reports described M. oleifera as highly potent anti-in Xammatory (Ezeamuzle et al., 1996), hepatoprotective (Pari and Kumar, 2002), antihypertensive (Faizi et al., 1995) and anti-tumor (Murakami et al., 1998). Also, its seed has strong coagulative and antimicrobial properties (Eilert et al., 1981). The seed oil has physical and chemical properties equivalent to that of olive oil and contains a large quantity of tocopherols (Tsaknis et al., 1999). The leaf extracts in rats were found to regulate thyroid status and cholesterol levels (Tahiliani and Kar, 2000; Ghasi et al., 2000). In recent years, many people in Taiwan or China have been using the seed of Moringa as an herbal medicine to treat athlete's foot and tinea and found that it is Vective. For the Wrest time, in this communication we provide the evidence that extracts of M. oleifera have anti-fungal properties. M. oleifera is a highly valued plant, distributed in many countries of the tropics and subtropics. It has an impressive range of medicinal uses with high nutrition value. Different parts of this plant contain a profile of important minerals, and are a good source of protein, vitamin, B carotene, amino acids, and various phenolics. In addition to its compelling water purifying powers and high nutritional value, M. oleifera is very important for its medicinal value. Various part of this plant such as the leaves, roots, seed, bark, fruit, flowers and immature pods acts as cardiac and circulatory stimulants, possess antitumor, antipyretic, antiepileptic, anti-inflammatory, antiulcer. antispasmodic, diuretic. antihypertensive. cholesterol lowering, antioxidant. antidiabetic. hepatoprotective, antibacterial and antifungal activities. They are being employed for the treatment of different ailments in the traditional system of medicine. This research work will focus on the detailed phytochemical composition, medicinal uses, along with pharmacological properties of different parts of this multipurpose tree (Dixit et al., 2016).

# 3. NEED FOR NOVEL APPROACH:

Antibiotic misuse is one of the main reasons of generating drug resistant pathogens. Patients who are non-compliant or who do not complete the course of antibiotic therapy also cause an increase in antibiotic-resistant bacteria. Multi drug resistant infecting human pathogens are a challenge for the clinicians across the globe. Plant materials used as herbal or ayurvedic drugs and raw materials for the pharmaceutical industries can be a promising alternative for chemical antibiotics due to quality, safety and efficacy. The pharmacological investigations of plants are being carried out to find novel drugs or templates for the development of new therapeutic agents to combat drug resistant pathogens (Iwu, 1999). The therapeutic applicability of silver and medicinal plants in treating bacterial infections is already well known (Gopinath et al., 2010; Chanda et al., 2013). Recently, synthesis of AgNPs with the help of medicinal plants is attempted; the reduction of silver to nano size is accomplished by the secondary metabolites present in the medicinal plants.

# 4. SYNTHESIS OF MORINGA OLIFERA SILVER NANOPARTICLES (MO-AGNPS)

Synthesis of nanoparticles (NPs) using plants can be advantageous over other biological processes by eliminating the elaborate process of maintaining cell culture (Willner, 2006). The microbial enzymes and secondary metabolites with antioxidant or reducing properties are usually reducing metal compounds into their respective NPs. Plants have been reported to be used for synthesis of metal NPs of gold and silver and of a goldsilver-copper alloy (Joerger et al., 2000; Ahmad et al., 2003, Anderson et al., 1998; Romero-Gonzlez et al., 2009; Gardea-Torresday et al., 2003). Colloidal silver is of particular interest because of its distinctive properties such as good conductivity, chemical stability, and catalytic and antibacterial activity (Tessier et al., 2000; Cao et al., 2002; Kumar et al., 2015). The first step of synthesis is to make aqueous extract, which is usually done by boiling the plant material in distilled water. This plant extract is added to silver nitrate (AgNO<sub>3</sub>) and the colour of AgNO<sub>3</sub> changes from colourless to yellow to brown to orange indicating the synthesis AgNPs in the aqueous solution. There are many factors which affect the formation of silver nano particles. The concentration of the aqueous plant extract plays an important role in the formation of AgNPs (Raman et al., After that AgNPs formed collected by centrifugation and dried AgNPs pallet used for further characterization (Poudel et al., 2017). The higher concentration of the plant extract will lead to the formation of more AgNPs. The biosynthesis of NPs, which represents a connection between biotechnology and nanotechnology, has received increasing consideration due to the growing need to develop environmentally friendly technologies for material syntheses. The search for appropriate biomaterials

for the biosynthesis of NPs continues through many different synthetic methods (Satishkumar et al., 2009)

# 5. CHARACTERIZATION OF MO-AGNPS

The synthesized AgNPs are generally characterized by UV-Vis spectroscopy, Scanning electron microscopy (SEM), Transmission electron microscopy (TEM), Fourier transform infrared spectroscopy (FTIR), Zeta potential and X-Ray diffraction measurement (XRD).

Ultraviolet-Visible (UV-VIS) spectroscopy: Ultravioletspectroscopy or ultraviolet-Visible spectrophotometer (UV-Vis) refers to absorption spectroscopy in the UV-Visible spectral region. This means it uses light in the visible and adjacent near-UV and nearinfrared (NIR) ranges. The absorption in the visible range directly affects the perceived color of the chemicals involved. In this region of the electromagnetic spectrum, molecules undergo electronic transitions (Das et al., 2013). It is well known that the optical absorption spectra of metal NPs are dominated by surface plasmon resonances (SPRs) that shift to longer wavelengths with increasing particle size. Also, it is well recognized that the absorbance of AgNPs depends mainly on the size and shape. In general, the number of SPR peaks decreases as the symmetry of the nanoparticle increases. The position and shape of the plasmon absorption depends on the particles size and shape, and the dielectric constant of the surrounding medium (Kelly et al., 2003, Rai et al, 2006). The appearance of SPR peaks at 446 nm provides a convenient spectroscopic signature for the formation of AgNPs (Basavaraja et al., 2008).

**Scanning electron microscopy (SEM):** The SEM analysis is employed to characterize the size, shape, morphology and distribution of synthesized AgNPs (Ramamurthy et al., 2013).

**Transmission electron microscopy** (**TEM**): TEM measurements are conducted in order to estimate the particle size and size distribution of the synthesized AgNPs (Zaved et al., 2012). The *Moringa oliefera* extract should be sufficient enough to be coated on the synthesized AgNPs, otherwise aggregation of particles is accelerated and the particles are not sufficiently stabilized.

**Fourier transform infrared spectroscopy (FTIR):** FTIR measurements are carried out to identify the possible biomolecules responsible for reduction, capping and efficient stabilization of AgNPs and the local molecular environment of the capping agents on the nanoparticles (Vidhu et al., 2011).

**Zeta potential:** Zeta potential is an essential parameter for the characterization of stability in aqueous nano suspension. A minimum of + 30 mV zeta potential values is required for indication of stable nano suspension (Jacobs and Muller, 2002) Higher zeta potential indicates greater stability of the synthesized AgNPs (Jebakumar and Sethuraman, 2012).

**X-Ray diffraction (XRD):** The XRD has proven to be a valuable research tool to prove the formation of AgNPs, and to determine the crystal structure of the prepared AgNPs and to calculate the crystalline particle size (Bindhu and Umadevi, 2013). Mounting evidences suggest that

AgNPs act as promising antimicrobial agents and may emerge as an alternative to conventional antibiotics. They could be of immense use in the medical field for their efficient antimicrobial function. The present review describes some of the most promising plants by the help of which AgNPs have been synthesized which can be used as a new novel source of antimicrobics to combat multiple drug resistant tough microorganisms and also can be therapeutically utilized to combat other diseases and disorders.

# 6. ANTIMICROBIAL ACTIVITY OF MO-AGNPS

The antibacterial activity exhibited by AgNPs depends on AgNO<sub>3</sub> concentration. It is inversely proportional i.e. less metal concentration more is the activity and vice versa. This is because smaller particles have larger surface area available for interaction and will give more bactericidal effect than the larger particles (Baker et al., 2005). The exact mechanism of the antibacterial effect of silver ions is not totally understood. Research reports reveal that the positive charge on the Ag ion is crucial for its antimicrobial activity. The antibacterial activity is probably derived, through the electrostatic attraction between negativelycharged cell membrane of microorganism and positivelycharged nanoparticles (Hamouda et al., 2000; Dibrov et al., 2002; Dragieva et al., 1999). NPs exhibit completely new or improved properties based on specific characteristics such as size, distribution and morphology. It was reported that the antimicrobial activity of AgNPs has strongly effective against the microorganisms in 100µl of AgNPs, owing to the surface of smaller size nanoparticles to change the local electronic structure for the enhancement of chemical reactivity to control the bactericidal effect (Mubarak Ali et al., 2011). The cell membrane of microorganisms is negatively charged and AgNPs are positively charged and when these positively charged silver nano particles accumulate on negatively charged cell membrane, it brings about a substantial conformational change in the membrane and it ultimately loses permeability control which leads to cell death (Ramamurthy et al., 2013; Hamouda and Baker, 2000). Mubarak Ali. et al. (Mubarak et al., 2011) stated that once AgNPs the bacterial cell, they would interfere with the bacterial growth signaling pathway by modulating tyrosine phosphorylation of putative peptides substrates critical for cell viability and cell division. Ag NPs adsorb the surface of bacteria but in low concentration does not enter the cells of bacteria; actually respiration occurs across the cell membrane other than mitochondrial membrane (Nabikhan et al., 2010). The NPs release silver ions in the bacterial cells, which enhance their bactericidal activity (Sondi and Salopek-Sondi, 2004; Marones et al., 2005). Mahendra et al. (2009) stated that AgNPs preferable attack the respiratory chain, cell division finally leading to cell death. According to Amro et al. (2000) metal depletion may cause the formation of irregularly shaped pits in the outer membrane and change membrane permeability, which is caused by progressive release of lipopolysaccharides and membrane proteins. Or perhaps DNA loses its replication ability and expression of ribosomal subunits proteins as

well as some other cellular proteins and enzymes essential to ATP production becomes inactivated (Sanghi and Verma, 2009). The other mechanism proposed by Danilczuk et al. (2006) and Kim et al. (2007) is the formation of free radicals which subsequently induces membrane damage leading to efficient antimicrobial property of AgNPs. The other mechanism proposed is involvement of interaction of AgNPs with biological macromolecules such as enzymes and DNA through an electro-release mechanism. The NPs get attached to the cell membrane and penetrate inside the bacteria. The bacterial membrane contains sulfur containing proteins and the silver nanoparticles interact with these proteins in the cell as well as with the phosphorus containing compounds like DNA. Their interaction may cause damage to DNA and proteins resulting in cell death. Ag+ binds to functional groups of proteins, resulting in protein denaturation (Guzman et al, 2012). The AgNPs show efficient antimicrobial property due to their extremely large surface area, which provides better contact with microorganisms. It is reasonable to state that the binding of the NPs to the bacteria depends on the interaction of the surface area available. Smaller particles having a larger surface area available for interaction will have a stronger bactericidal effect than will larger particles (Shrivastava et al, 2007; Guzman et al, 2012).

# 7. APPLICATIONS OF MO-AGNPS:

Antimicrobial capability of AgNPs allows them to be suitably employed in numerous household products such as textiles, food storage containers, home appliances and in medical devices. The most important application of silver and AgNPs is in medical industry such as tropical ointments to prevent infection against burn and open Synthesized MO-AgNPs showed significant topical antifungal activity related to decreased particle size and increased surface area. Vibhute et al reported that both topical formulations viz. cream and ointment found to be stable after accelerated stability study (Vibhute et al. 2014). Numerous pharmacological investigations of M. oleifera have been reported on anti-inflammation, anti-infection, antidiabetic, antioxidant, and antihyperlipidemic activities (Singh and Singhet, 2009; Ndong et al., 2007; Verma et al., 2009; Chumark et al. 2008; Vongsak et al. 2012; Jung et al., 2010). Recently, isoquercetin, astragalin, and cryptochlorogenic acid were reported to be major active components in M. oleifera (Fernandez et al 2005). Isoquercetin is a powerful natural antioxidant which possesses several potential therapeutic effects including antiasthma and antihypertension (Gasparotto et al., 2011; Park et al., 2012, Soromou et al., 2012). Astragalin is also reported as a natural antioxidant agent exhibiting some biological properties such as attenuation of inflammation, inhibition of dermatitis, and cellular protective effect (Kotani et al., 2000; Nakatani et al., 2000). Chlorogenic acid and its isomers are esters of quinic and caffeic acids that have abilities to inhibit oxidation and also promote various pharmacological activities such as antiobesity, reduction of plasma and liver lipids, and inhibition of acute lung injury (Cho et al, 2010). AgNPs are reported to have many therapeutic uses. There are reported to possess antiviral (Elechiguerra et al., 2005), antibacterial (Duran et al., 2005; Lee et al. 2003), antifungal (Krishnaraj et al., 2012), anti-parasitic (Santosh et al., 2012; Zahir and Rahuman, 2012), larvicidal activity (Jayaseelan et al., 2011; Rajakumar and Rahuman, 2011) and anticancer (Sukirtha et al., 2012; Gengan et al., 2013) properties. Due to strong antibacterial property AgNPs are used in clothing, food industry, sunscreens, cosmetics and many household appliances (Wijnhoven et al, 2009). Few studies have showed that AgNPs kill fungal spores by destructing the membrane integrity (Das et al 2013). NPs have numerous other benefits as well. Green synthesized AgNPs were also used for impregnation of polymeric medical devices to increase their antibacterial activity. Silver impregnated medical devices like surgical masks and implantable devices showed significant antimicrobial efficiency (Furno, 2007). There are reports that explored use of silver ions or metallic silver as well as AgNPs in medicine for burn treatment, dental materials, coating stainless steel materials, textile fabrics, water treatment, sunscreen lotions (Duran et al, 2007; Virk et al., 2019).

# 8. CONCLUSION

In conclusion, it has been demonstrated that the aqueous extract of *Moringa oleifera* leaf are capable of producing AgNPs extracellularly and the AgNPs are quite stable in solution. The formed AgNPs showed considerable antimicrobial activity compared to the respective antibiotics. The biosynthesis AgNPs prove to be potential candidates for medical application where antimicrobial activity is essential. Antibiotic resistance by the pathogenic bacteria has been observed since last decade; hence, the researchers are focusing on the development of new antibacterial agents which can overcome such resistance. Hence, MO-AgNPs proves to be an important step in this direction.

# **REFERENCES:**

- Agarwal A, Gayathri M (2017). Biological synthesis of nanoparticles from medicinal plants and its uses in inhibiting biofilm formation. Asian Journal of Pharmaceutical and clinical research, 10(5): 64-68
- Ahmad A, Mukherjee P, Senapati S, Mandal D, Khan MI, Kumar R, Sastry M. (2003). Extracellular biosynthesis of silver nanoparticles using the fungus *Fusarium oxysporum*. Colloids Surf., 28: 313.
- Ahmad A, Senapati S, Khan MI, Kumar R, Sastry M (2003). Extracellular biosynthesis of monodisperse gold nanoparticles by a novel extremophilic actinomycete thermomonospora sp. *Langmuir*, 19: 3550-3553.
- Amro NA, Kotra LP, Wadu-Mesthrige K, Bulychev A, Mobashery S, Liu G (2000). High-resolution atomic force microscopy studies of the *Escherichia coli* outer membrane: structural basis for permeability. *Langmuir*, 16: 2789-2796.
- Anderson CWN, Brooks RR, Stewart RB, Simcock R (1998). Harvesting a crop of gold in plants. *Nature*, 395: 553-554.
- Baker C, Pradhan A, Pakstis L, Darrin JP, Ismat SS (2005). Journal of Nanoscience Nanotechnology, 5: 244-249.
- Basavaraja S, Balaji SD, Lagashetty A, Rajasab AH, Venkataraman A (2008). Extracellular biosynthesis of silver nanoparticles using the fungus Fusarium semitectum. Material Research Bulletin. 43:1164-1170.
- Bhusnure OG, Jadhav PP, Hindole SS, Gholve SB, Giram PS, Kuthar VS (2017). Green Synthesis of Silver Nanoparticle Using Moringa oleifera Lam extract for pharmacological Activity. International Journal of Pharmacy and Pharmaceutical Research, 10(4): 59-66.

- Bindhu MR, Umadevi M (2013). Synthesis of monodispersed silver nanoparticles using *Hibiscus cannabinus* leaf extract and its antimicrobial activity. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy. 101: 184-190.
- Burali SC, Kanagralkar V, Sravani OS, Patil SL (2010). The beneficial effect of ethanolic extract of *Moringa oleifera* on osteoporosis. International Journal of Pharmaceutical Applications, 1(1): 50-58.
- Cao YC, Jin R, Mirkin CA (2002). Nanoparticles with Raman spectroscopic fingerprints for DNA and RNA detection. *Science*, 297: 1536-1540.
- Chanda S, Rakholiya K, Dholakia K, Baravalia Y (2013).
   Antimicrobial, antioxidant and synergistic property of two nutraceutical plants: *Terminalia catappa* L. and *Colocasia esculentum* L. *Turkish Journal of Biology*, 37: 81-91.
- Cho AS, Jeon, S M, Kim MJ (2010). Chlorogenic acid exhibits antiobesity property and improves lipid metabolism in high-fat dietinduced-obese mice. Food and Chemical Toxicology, 48 (3): 937– 943
- Chuang PH, Lee CW, Chou JY, Murugan M, Shieh BJ, Chen HM (2007). Anti-fungal activity of crude extracts and essential oil of *Moringa oleifera* Lam. Biosource Technology, 98: 232-236.
- Chumark P, Khunawat P, Sanvarinda Y (2008). The *in vitro* and *ex vivo* antioxidant properties, hypolipidaemic and antiatherosclerotic activities of water extract of *Moringa oleifera* Lam. Leaves. *Journal of Ethnopharmacology*, 116 (3), 439-446.
- Danilczuk M, Lund A, Saldo J, Yamada H, Michalik J (2006).
   Conduction electron spin resonance of small silver particles. Spectrochimaca Acta Part A, 63: 189-191.
- Das S., Parida UK, Bindhani BK (2013). Green Biosynthesis of Silver nanoparticles using Moringa oliefera. International Journal of nanotechnology and application, 3(2): 51-62.
- Dibrov P, Dzioba J, Gosink KK, Hase CC (2002). Chemiosmotic mechanism of antimicrobial activity of Ag(+) in Vibrio cholera, Antimicrob. Agents Chemother, 46: 2668.
- Dixit S., Tripathi A, Kumar P (2016). Medicinal Properties of Moringa oliefera- A review. International Journal of education and Science research review 3(2): 173-185.
- Dragieva I, Stoeva S, Stoimenov P, Pavlikianov E, Klabunde K (1999). Extracelluar sysnthesis of silver nanoparticles using dried leaves. *Nanostruct. Mater.*, 12: 267-72.
- Duran N, Marcato PD, Alves OL, Souza GI, Esposito E (2005). Mechanistic aspects of biosynthesis of silver nanoparticles by several Fusarium oxysporum strains. Journal of Nanobiotechnology, 13: 3-8.
- Duran N, Marcato PD, Souza GI, Alves OL, Esposito E (2007).
   Antibacterial effect of silver nanoparticles produced by fungal process on textile fabrics and their effluent treatment. *Journal of Nanobiotechnology*, 3: 203-208.
- Eilert U, Wolters B, Nahrstedt A (1981). The antibiotic principle of seeds of *Moringa oleifera* and *Moringa stenopetala*. *Planta Med.*, 42(1): 55-61.
- 24. Elechiguerra JL, Burt JL, Morones JR, Camacho BA, Gao X, Lara HH, Yacaman MJ (2005). Interaction of silver nanoparticles with HIV-1. *Journal of Nanobiotechnology*, 3: 6.
- Ezeamuzle IC, Ambadederomo AW, Shode FO, Ekwebelem SC (1996). Antiin Xammatory eVects of *Moringa oleifera* root extract. *International Journal of Pharmacognosy*, 34: 207-212.
- Fahey JW (2005). Moringa oleifera: A review of the medical evidence for its nutritional, therapeutic and prophylactic properties Part-1. Trees for life journal, 1(5): 1-15.
- Faizi S, Siddiqui BS, Saleem R, Siddiqui S, Aftab K, Gilani AH (1995). Fully acetylated carbonate and hypotensive thiocarbamate glycosides from *Moringa oleifera*. *Phytochemistry*, 38: 957-963.
- Fernandez J, Reyes R, Ponce H (2005). Isoquercitrin from Argemone platyceras inhibits carbachol and leukotriene D4- induced contraction in guinea-pig airways. European Journal of Pharmacology, 522 (1-3): 108-115.
- Fuhrman B (2005). Pomegranate Juice Inhibits Oxidized LDL Uptake and Cholesterol Bio- synthesis in Macrophages. *The Journal of Nutritional Biochemistry*, 16(9): 570-576.
- Furno F (2007). Silver nanoparticles and polymeric medical devices: a new approach to prevention of infection. *J Antimicrob Chemother*, 54:1019-1024.

- Gardea-Torresdey JLE, Gomez E, Peralta-Videa JR, Parsons JG, Troiani H, JoseYacaman M (2003). Alfalfa sprouts: a natural source for the synthesis of silver nanoparticles. *Langmuir.*, 2003; 19: 1357-1361.
- Gasparotto A, Gasparotto FM, Lourenco ELB (2011).
   Antihypertensive effects of isoquercitrin and extracts from Tropaeolum majus L.: evidence for the inhibition of angiotensin converting enzyme. Journal of Ethnopharmacology, 134 (2): 363-372
- Gengan RM, Anand K, Phulukdaree A, Chuturgoon A (2013). A549 lung cell line activity of biosynthesized silver nanoparticles using Albizia adianthifolia leaf. Colloids and Surfaces B. Biointerfaces, 105: 87-91.
- Ghasi S, Nwobobo E, Owli JO (2000). Hypocholesterolemic eVects of crude extract of leaf of *Moringa oleifera* Lam in high-fat diet fed Wistar rats. *Journal of Ethnopharmacology*, 69: 21-25.
- Gopinath P, Gogoi SK, Sanpoi P, Paul A (2010). Chattopadhyay, Ghosh SS. Signaling gene cascade in silver nanoparticle induced apoptosis. Colloids and Surfaces B: Biointerfaces, 77: 240-245.
- Gosh N, Paul S, Basak P 2014. Silver nanoparticles of Moringa oleifera- green synthesis, characterization and its antimicrobial efficacy. Journal of Drug Delivery and Therapeutics: Special Issue, 1: 42-46.
- 37. Gupta A, Gautam MK, Singh RK, Vijay Kumar M, Rao CV, Goel RK, Anuparba S (2010). Immunomodulatory effect of *Moringa oleifera* Lam extract on cyclophospamide induced toxicity in mice. *Indian Journal of Experimental Biology*, (48): 1157-1160.
- 38. Guzman M, Dille J, Godet S (2012). Synthesis and antibacterial activity of silver nano particles against gram positive and gram negative bacteria. *Nanomedicine:Nanotechnology, Biology and Medicine*, 8:37-45.
- Halliwell B (1997). Antioxidants and human disease: a general Introduction. Nutr Rev., 55: S44

  –S52.
- Hamouda T, Baker Jr JR (2000). Antimicrobial mechanism of action of surfactant lipid preparations in enteric gram-negative bacilli. *Journal of Applied Microbiology*, 89: 397-403.
- Hamouda T, Myc A, Donovan B, Shih A, Reuter JD, Baker Jr (2000). A novel surfactant nanoemulsion with a unique non-irritant topical antimicrobial activity against bacteria, enveloped viruses and fungi, J. R. Microbiol. Res., 56: 1.
- Iwu MW (1999). New antimicrobials of plant origin. In: J. Janick (ed.), Perspectives on new crops and new uses. ASHS Press, Alexandria, VA, pp. 457-462.
- 43. Jacobs C, Müller RH (2002). Production and characterization of a budesonide anosuspension for pulmonary administration. *Pharmaceutical Research.* 19: 189-194.
- Jaiswal D, Rai PK, Kumar A, Mehta S, Watal G (2009). Effects of Moringa oleifera Lam, leaves aqueous extract therapy on hyperglycemic rats. Journal of Ethnopharmacology, 123: 392-396.
- 45. Jayaseelan C, Rahuman AA, Rajakumar G, Kirthi AV, Santhoshkumar T, Marimuthu S, Bagavan A, Kamaraj C, Zahir AA, Elango G (2011). Synthesis of pediculocidal and larvicidal silver nanoparticles by leaf extract from heartleaf moonseed plant, *Tinospora cordifolia* Miers. *Parasitology Research*, 109: 185-194.
- Jebakumar IET, Sethuraman MG (2012). Instant green synthesis of silver nanoparticles using *Terminalia chebula* fruit extract and evaluation of their catalytic activity on reduction of methylene blue. *Process Biochemistry*. 47: 1351-1357.
- 47. Jeong SH, Yeo SY, Yi SC (2005). The effect of filler particle size on the antibacterial properties of compounded polymer/silver fibers. *J. Mat. Sci.*, 40: 5407-5411.
- Joerger R., Klaus T. and Granqvist C.G. (2000). Biologically produced silver-carbon composite materials for optically functional thin-film coatings. Advanced Materials., 12: 407-409.
- Jung SH, Kim BJ, Lee EH, Osborne NN (2010). Isoquercitrin is the most effective antioxidant in the plant *Thuja orientalis* and able to counteract oxidative-induced damage to a transformed cell line (RGC-5 cells). *Neurochemistry International*, 57 (7): 713-721.
- Kelly KL, Coronado E, Zhao LL, Schatz GC (2003). The optical properties of metal nano particles: The influence of size, shape and dielectric environment. *Journal of Physical Chemistry*, 107: 668-677.
- Kim JS, DVM, Kuk E, Yu KN, Kim J-H, Jin Park SJ, Lee HJ, Kim SH, Young Kyung Park YK, Park YH, Hwang C-Y, Kim YK, Lee YS, Jeong DH, Cho MH (2007). Antimicrobial effects of silver

- nanoparticles. Nanomedicine: Nanotechnology, Biology, and Medicine, 3: 95-101.
- Kotani M, Matsumoto M, Fujita A (2000). Persimmon leaf extract and astragalin inhibit development of dermatitis and IgE elevation in NC/NGa mice. *Journal of Allergy and Clinical Immunology*, 106 (1): 159-166.
- Krishnaraj C, Ramachandran R, Mohan K, Kalaichelvan PT (2012).
   Optimization for rapid synthesis of silver nanoparticles and its effect on phytopathogenic fungi. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy. 93: 95-99.
- Kumar RSDS, Kumar LB, Kumar SP, Chandirasekar R, Kumar UV (2015). Biomimetics of silver nanoparticles from *Ganoderma lucidum* (Curtis) P Karst and its anticancer potential on breast cancer cells. *Int J Recent Adv Multidiscipline Res*, 2(11): 903-909.
- Lee S, Yeo HJ, Jeong SY (2003). Antibacterial effect of nanosized silver colloidal solution on textile fabrics. *Journal of Material* Science, 38: 2199-2204.
- Liu RH (2002). Supplement quick fix fails to deliver. Food Technol Int., 1: 71-72.
- Mahendra R, Alka Y, Aniket G (2009). Silver nanoparticles as a newgeneration of antimicrobials. Biotechnology Advances, 27: 76-83
- Mohamed MB, Volkov V, Link S, Sayed MAE (2000). The 'lightning' gold nanorods: fluorescence enhancement of over a million compared to the gold metal. *Chem Phys Lett.*, 317: 517-523.
- Mohanpuria P, Rana NK, Yadav SK (2008). Biosynthesis of nanoparticles: technological concepts and future applications. *Journal Nanopart Res.*, 10: 507-517.
- Morones JR, Elechiguerra JL, Camacho A, Holt K, Kouri JB, Ramfrez JT, Yacaman MJ (2005). The bactericidal effect of silver nanoparticles. *Nanotechnology*, 16: 2346-2353.
- Mubarak Ali D, Thajuddin N, Jeganathan K, Gunasekaran M (2011).
   Plant extract mediated synthesis of silver and gold nanoparticles and its antibacterial activity against clinically isolated pathogens. Colloids and Surfaces B: Biointerfaces, 85: 360-365.
- Murakami A, Kitazono Y, Jiwajinda S, Koshimizu K, Ohigashi H (1998). Niaziminin, a thiocarbamate from the leaves of *Moringa oleifera*, holds a strict structural requirement for inhibition of tumorpromoter- induced Epstein–Barr virus activation. *Planta Medica*, 64: 319-323.
- Nabikhan A, Kandasamy K, Raj A, Alikunhi NM (2010). Synthesis of antimicrobial silver nanoparticles by callus and leaf extracts from salt marsh plant, Sesuvium portulacastrum L. Collo. Surf. B: Biointerfaces, 79: 488-493.
- Nakatani N, Kayano SI, Kikuzaki H, Sumino K, Katagiri K, Mitani T (2000). Identification, quantitative determination, and antioxidative activities of chlorogenic acid isomers in prune (*Prunus domestica L.*). *Journal of Agricultural and Food Chemistry*, 48(11): 5512-5516.
- Ndong M, Uehara M, Katsumata SI, Suzuki K (2007). Effects of oral administration of *Moringa oleifera* Lam on glucose tolerance in Goto-Kakizaki and wistar rats. *Journal of Clinical Biochemistry and* Nutrition, 40(3): 229-233.
- Pari L, Kumar NA (2002). Hepatoprotective activity of *Moringa oleifera* on antitubercular drug-induced liver damage in rats. *Journal of Medicinal Food*, 2002; 5(3): 171-177.
- Park SN, Kim SY, Lim GN, Jo NR, Lee MH (2012). In vitro skin permeation and cellular protective effects of flavonoids isolated from Suaeda asparagoides extracts. Journal of Industrial and Engineering Chemistry, 18(2): 680-683.
- Patel RK, Patel MM, Patel MP, Kanzaria NR, Vaghela KR, Patel NJ (2008). Hepatoprotective Activity of *Moringa oleifera* Lam. fruit on isolated rat hepatocytes. *Pharmacognosy Magazine*, 4(15): 118-123.
- Poudel M, Pokharel R, Sudip KC, Awal SC, Pradhananga R (2017). Biosynthesis of Silver Nanoparticles Using *Ganoderma lucidum* and Assessment of Antioxidant and Antibacterial Activity. *Int J Appl Sci Biotechnol*, 5(4): 523-531.
- Prabhu N, Divya TR, Yamuna G (2010). Synthesis of silver phyto nanoparticles and their antibacterial efficacy. *Digest.J. Nanomater. Biostruct.* 185-189.
- Prakash A (1998). Ovarian response to aqueous extract of Moringa oleifera. Fitoterapia, 59: 89-91.
- Rai A, Singh A, Ahmad A, Sastry M (2006). Role of halide ions and temperature on the morphology of biologically synthesized gold nanotriangles. *Langmuir*. 22: 736-741.

- Rai M, Yadav A, Gade A (2009). Silver nanoparticles as a new generation of antimicrobials. *Biotechnology Advances*, 27: 76-83.
- Rajakumar G, Rahuman AA (2011). Larvicidal activity of synthesized silver nanoparticles using *Eclipta prostrata* leaf extract against filariasis and malaria vectors. *Acta Tropica*. 118: 196-203.
- Ramamurthy CH, Padma M, I. Samadanam DM, Mareeswaran R, Uyavaran A, Suresh Kumar M, Premkumar K, Thirunavukkarasu C (2013). The extra cellular synthesis of gold and silver nanoparticles and their free radical scavenging and antibacterial properties. Colloids and Surfaces B: Biointerfaces, 102: 802-815.
- Raman N, Sudharsan S, Veerakumar V, Pravin N, Vithiya K (2012). Pithecellobium dulce mediated extra-cellular green synthesis of larvicidal silver nanoparticles. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, 96: 1031-1037.
- Rastogi T, Bhutda V, Moon K, Aswar PB, Khadabadi SS (2009).
   Comparative Studies on Anthelmintic Activity of Moringa oleifera & Vitex negund. Asian Journal of Research in Chemistry, 2(2): 181-182.
- Rathi BS, Bodhank SL, Baheti AM (2006). Evaluation of aqueous leaves extract of *Moringa oleifera* Linn. for wound healing in albino rats. *Indian Journal of Experimental Biology*, 44: 898-901.
- Romero-Gonzalez J, Walton JC, Peralta-Videa JR, Rodriguez E, Romero J, GardeaTorresdey JL (2009). Modeling the adsorption of Cr(III) from aqueous solution onto Agave lechuguilla biomass: study of the advective and dispersive transport. *Journal of Hazardous Materials*, 161: 360-365.
- Sanghi R, Verma P (2009) . Biomimetic synthesis and characterization of protein capped silver nanoparticles. *Bioresource Technology*, 100: 501-504.
- 81. Santhosh kumar T, Rahuman AA, Bagavan A, Marimuthu S, Jayaseelan C, Vishnu Kirthi A, Kamaraj C, Rajakumar G Zahir AA, Elango G, Velayutham K, Iyappan M, Siva C, Karthik L, Rao VBK (2012). Evaluation of stem aqueous extract and synthesized silver nanoparticles using *Cissus quadrangularis* against *Hippobosca maculata* and *Rhipicephalus* (Boophilus) *microplus*. *Experimental Parasitology*. 132: 156-165.
- Sathishkumar M, Sneha K, Won SW, Cho CW, Kim S, Yun YS (2009). Cinnamon zeylanicum bark extract and powder mediated green synthesis of nano-crystalline silver particles and its bactericidal activity. Colloid Surface B., 73: 332-338.
- Shahverdi AR, Minaeian S, Shahverdi HR, Jamalifar H, Nohi AA (2007). Rapid synthesis of silver nanoparticles using culture supernatants of Enterobacteria: A novel biological approach. *Proc. Biochem.* 2: 919.
- 84. Shrivastava S, Bera T, Roy A, Singh G, Ramachandrarao P, Dash D (2007). Characterization of enhanced antibacterial effects of novel silver nanoparticles. Nanotechnology, 18: 225103.
- Shukla S, Mathur R, Prakash AO (1981). Effects of aqueous extract of *Moringa oleifera* Lam. on the periodicity of oestrous cycle in adult intact rats. *Indian Journal of Pharmacological Science*, 49: 218-219.
- 86. Siddhartha D, Guha D (2007). Role of *Moringa oleifera* on enterochromaffin cell count and serotonin content of experimental ulcer model. *Indian Journal of Experimental Biology*, 726-731.
- 87. Silva NC, Júnior AF (2010). Biological properties of medicinal plants: A review of their antimicrobial activity. *JVenom Anim Toxins Incl Trop Dis*, 16(3): 402-13.
- Singh BN, Singhet RL (2009). Oxidative DNA damage protective activity, antioxidant and anti-quorum sensing potentials of *Moringa* oleifera. Food and Chemical Toxicology, 47(6): 1109–1116.
- Sondi I, Salopek-Sondi B (2004). Silver nanoparticles as antimicrobial agent: a case study on E. coli as a model for Gramnegative bacteria. Journal of Colloids Interface Science. 275: 177-182
- Soromou LW, Chen N, Jiang L (2012). Astragalin attenuates lipopolysaccharide-induced inflammatory responses by downregulating NF-\_B signaling pathway. *Biochemical and Biophysical Research Communications*, 419 (2): 256-261.
- Sukirtha R, Priyanka KM, Antony JJ, Kamalakkannan S, Thangam R, Gunasekaran P, Krishnan M, Achiraman S (2012). Cytotoxic effect of Green synthesized silver nanoparticles using *Melia* azedarach against in vitro HeLa cell lines and lymphoma mice model. Process Biochemistry, 47: 273-279.

- Sumner MD (2005). Effects of Pomegranate Juice Consumption on Myocardial Perfusion in Patients with Coronary Heart Disease. American Journal of Cardiology, 96(6): 810-814.
- 93. Tahiliani P, Kar A 2000. Role of *Moringa oleifera* leaf extract in regulation of thyroid hormone status in adult male and female rats. *Pharmacological Research*, 41: 319-323.
- Tessier PM, Velev OD, Kalambur AT, Rabolt JF, Lenhoff AM, Kaler EW (2000). Assembly of gold nanostructured films templated by colloidal crystals and use in surface enhanced Raman spectroscopy. *Journal of the American Chemical Society*, 122: 9554-9555.
- Tsaknis J, Lalas S, Gergis V, Dourtoglou V, Spilotis V (2009). Characterisation of Moringa oleifera variety Mbololo seed oil of Kenya. Journal of Agricultural and Food Chemistry, 47: 4495-4499.
- Verma AR, Vijayakumar M, Mathela CS, Rao CV (2009). In vitro and in vivo antioxidant properties of different fractions of *Moringa* oleifera leaves. Food and Chemical Toxicology, 47(9): 2196-2201.
- Vibhute SK, Kasture VS, Kendra PN, Wagh GS 2014. Synthesis of Silver nanoparticles from *Moringa oleifera*: Formulation and evaluation against *Candida albicans*. Indo American journal of Pharmaceutical Research, 4(3): 1581-1587.
- Vidhu VK, Aromal SA, Philip D (2011). Green synthesis of silver nanoparticles using Macrotyloma uniflorum. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, 83: 392-397.

- Virk AK, Kulshrestha S, Thakur C, Tripathi A, Kakade A, Li X (2018). Development and efficacy analysis of a Moringa oliefera based potable water purification kit. Journal of Water Process Engineering, 27: 37-64.
- Vongsak B, Sithisarn P, Gritsanapan W (2012). HPLC quantitative analysis of three major antioxidative components of *Moringa* oleifera leaf extracts. *Planta Medica*, 78(11): 1252.
- 101. Wijnhoven SWP, Peijnenburg WJGM, Herberts CA, Hagens WI, Oomen AG, Heugens EHW, Roszek B, Bisschops J, Gosens I, Van de Meent D, Dekkers S, de Jong WH, Van Zijverden M, Sips AJAM, Geertsma RE (2009). Nano-silver: a review of available data and knowledge gaps in human and environmental risk assessment. Nanotoxicology, 3: 109.
- Willner I, Baron R, Willner B (2006) Growing metal nanoparticles by enzymes. Adv Mater., 18: 1109-1120.
- 103. Zahir AA, Rahuman AA (2012). Evaluation of different extracts and synthesised silver nanoparticles from leaves of Euphorbia prostrata against Haemaphysalis bispinosa and Hippobosca maculate. Veterinary Parasitology, 187: 511-520.
- 104. Zaved MF, Eisa WH, Shabaka AA (2012). *Malva parviflora* extract assisted green synthesis of silver nanoparticles. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 98: 423-428.